# **Molecular Programming**

# Luca Cardelli

**Microsoft Research** 

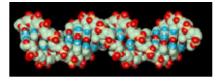
Redmond, 2009-06-29 http://LucaCardelli.name

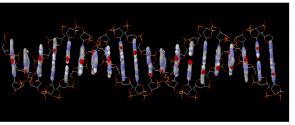
# **DNA Basics**

### ACGT

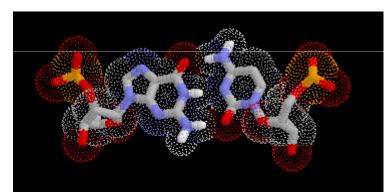
#### **Interactive DNA Tutorial**

(http://www.biosciences.bham.ac.uk/labs/minchin/tutorials/dna.html)

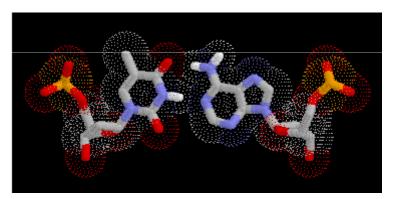




Sequence of Base Pairs



GC Base Pair Guanine-Cytosine

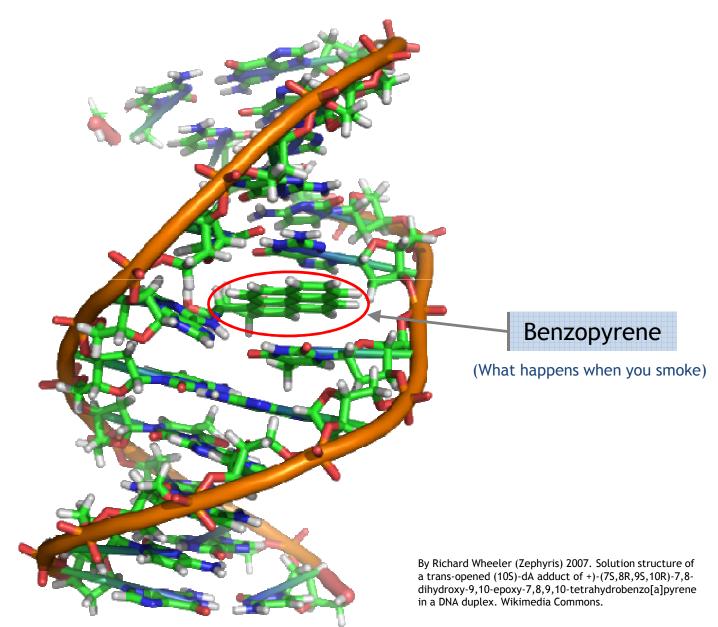


TA Base Pair Thymine-Adenine

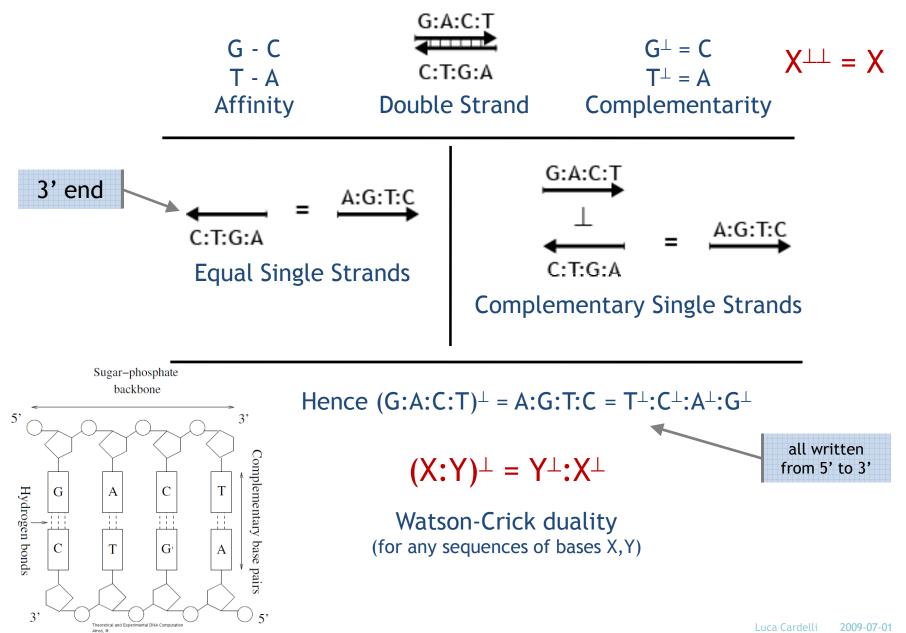
Hence DNA is a string over a 4-letter ACGT alphabet

Human genome : ~3 billion base pairs = 750 Megabytes (since 1 byte encodes 4 base pairs) = 1 movie download!

### **DNA Double Helix**

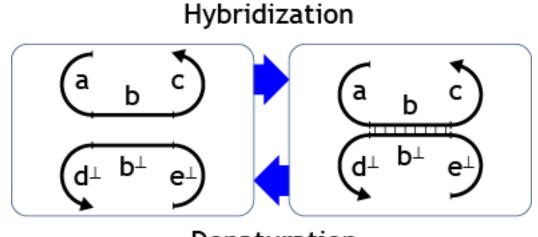


## Watson-Crick Duality



ental DNA Computatio

### **Hybridization**



Denaturation

Hybridization is also called annealing; denaturation is also called melting.

The direction of the reaction (or in general the equilibrium between the two states) is determined by a number of factors, e.g. temperature.

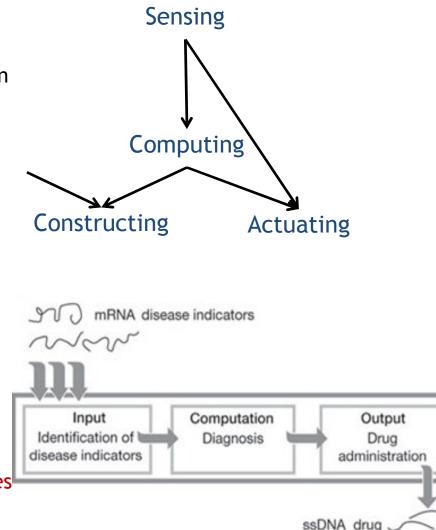
We assume we are in conditions that favor hybridization beyond a certain length of matching region.

# **DNA Nanotechonology**

## Nano Tasks

#### • Sensing

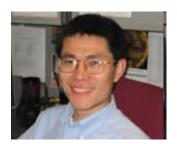
- Binding to specific molecules
- Computing
  - Analog: Signal Filtering or Amplification
  - Digital: Logical gates
- Actuating
  - Releasing molecules
  - Producing forces
- Constructing
  - $\circ$  By self-assembly
  - $\circ$  Or under 'program' control
- Nucleic Acids (DNA/RNA)
  - Probably the only materials that can perform all these functions.
  - Technology relatively well developed.
  - They can interface to biological entities



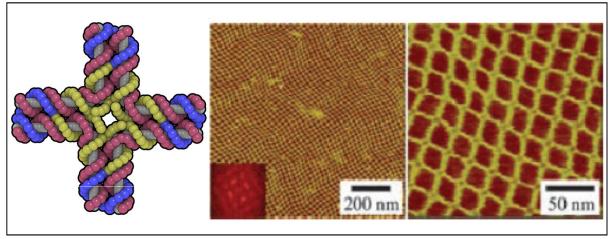
# **DNA as a Building Material**

Slides by John Reif

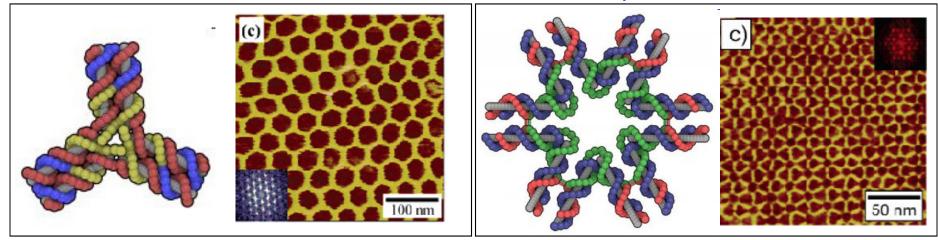
# **2D DNA Lattices**

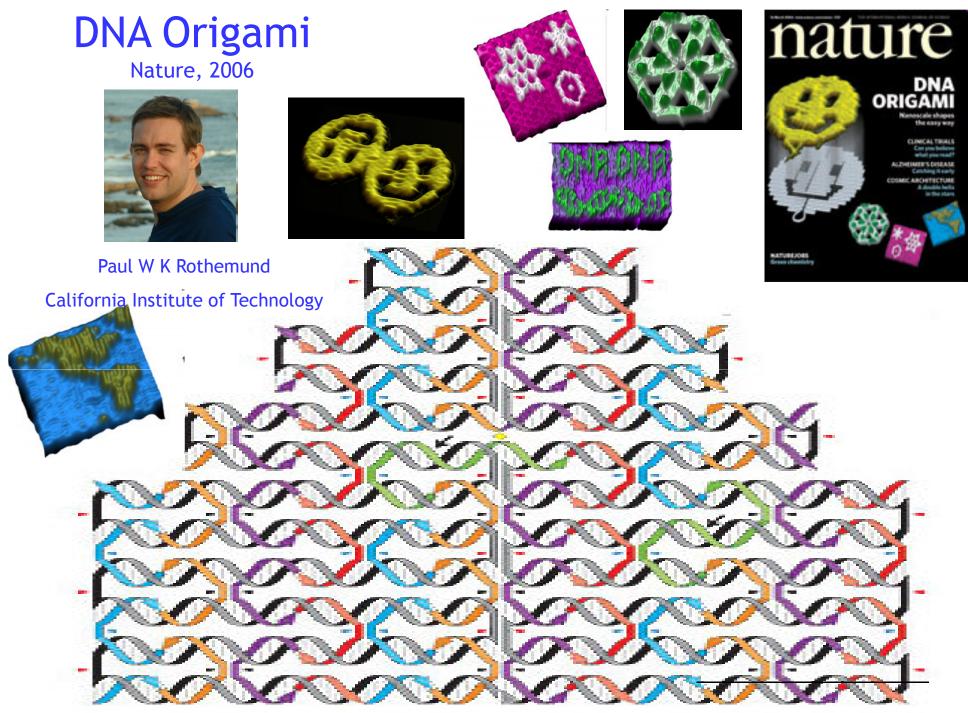


Chengde Mao Purdue University, USA

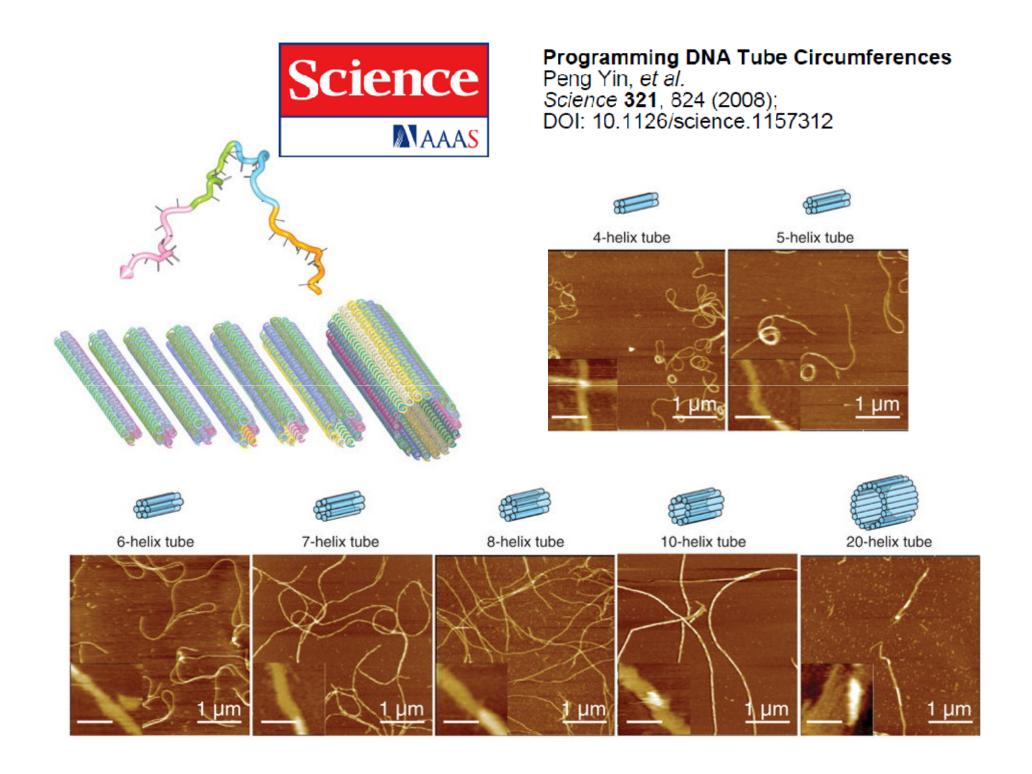


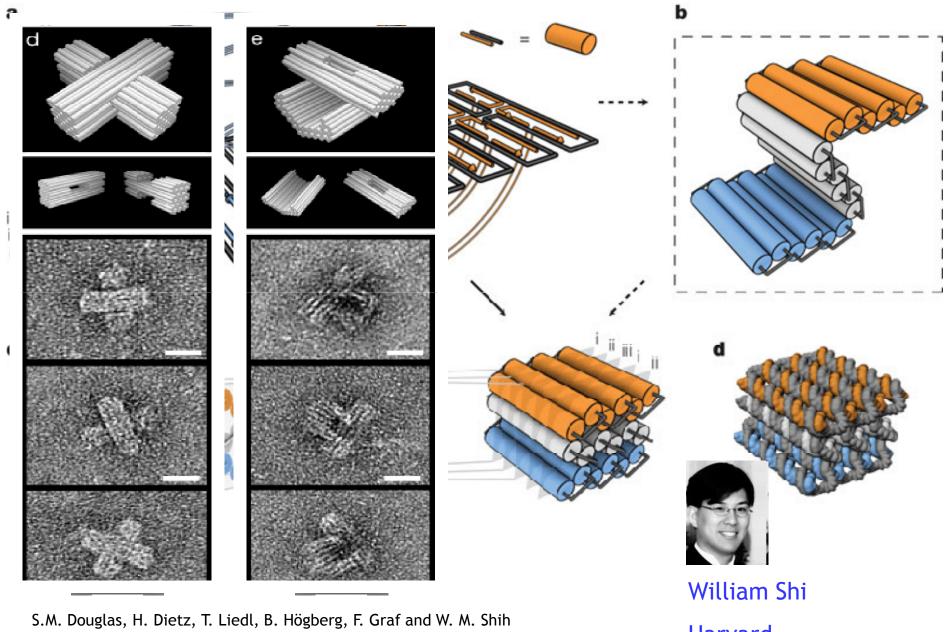
#### N-point Stars





PWK Rothemund, *Nature* 440, 297 (2006)

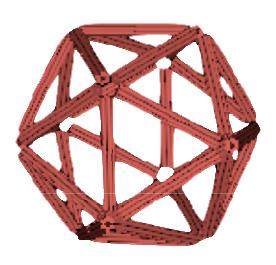


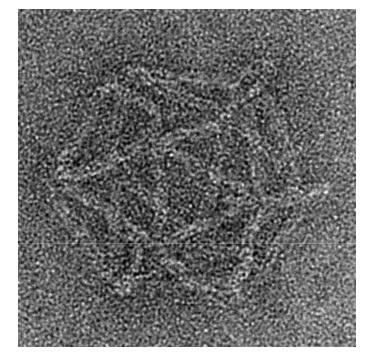


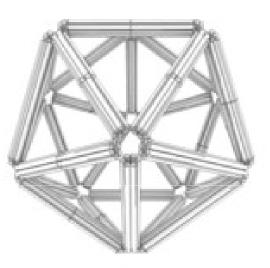
Self-assembly of DNA into nanoscale three-dimensional shapes, Nature (2009)

Harvard

#### 3D Wireframe Icosahedron









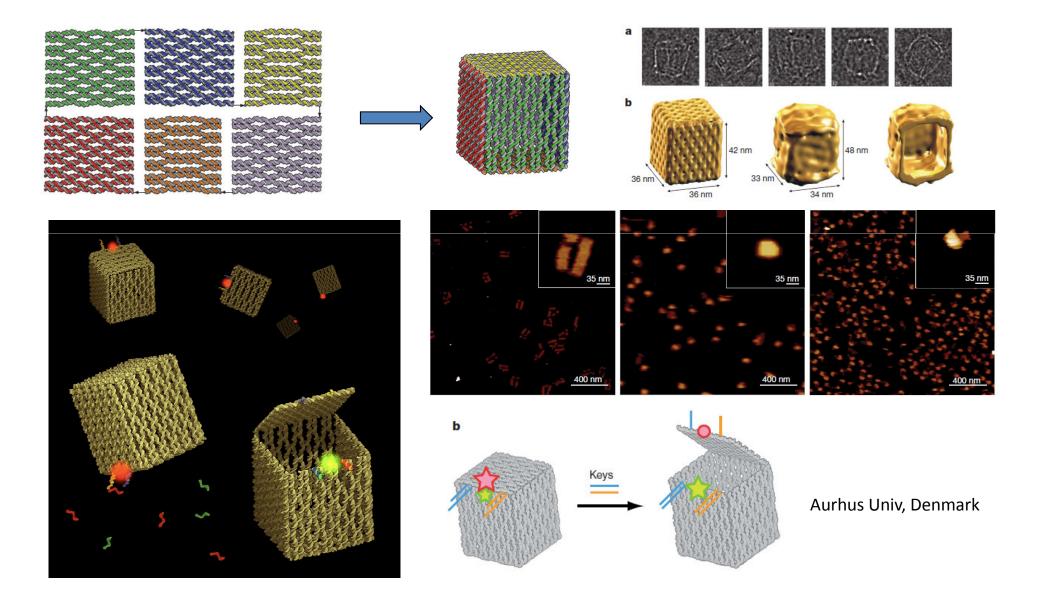
#### William Shi

#### Harvard

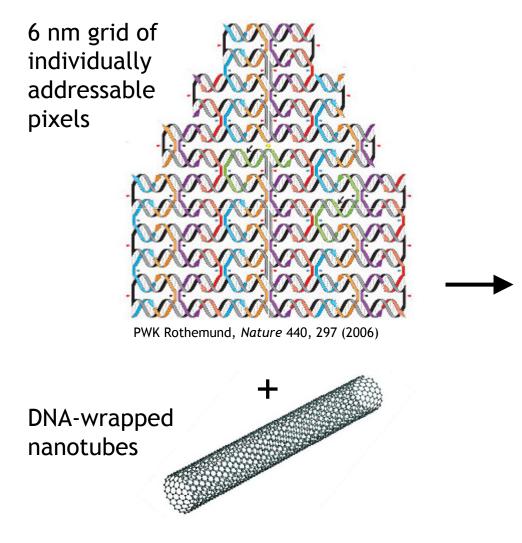
S.M. Douglas, H. Dietz, T. Liedl, B. Högberg, F. Graf and W. M. Shih Self-assembly of DNA into nanoscale three-dimensional shapes, Nature (2009)

## Self-assembly of a DNA origami box

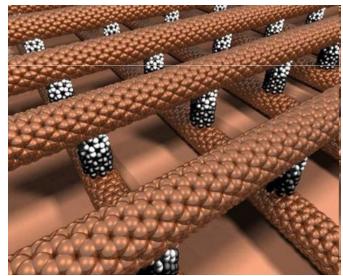
Andersen et al *Nature* **2009**, *459*, 73



## DNA circuit boards (IBM )



"What we are really making are tiny DNA circuit boards that will be used to assemble other components." --Greg Wallraff, IBM

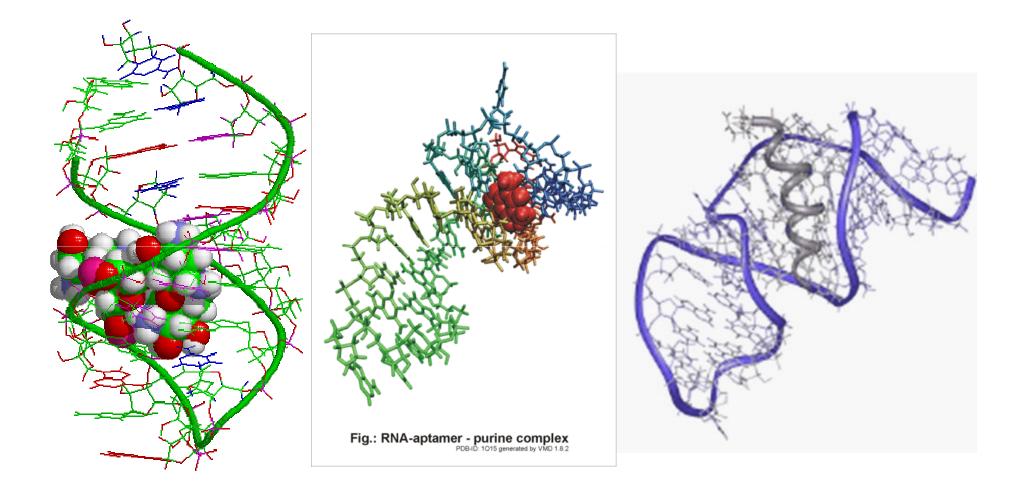


European Nanoelectronics Initiative Advisory Council

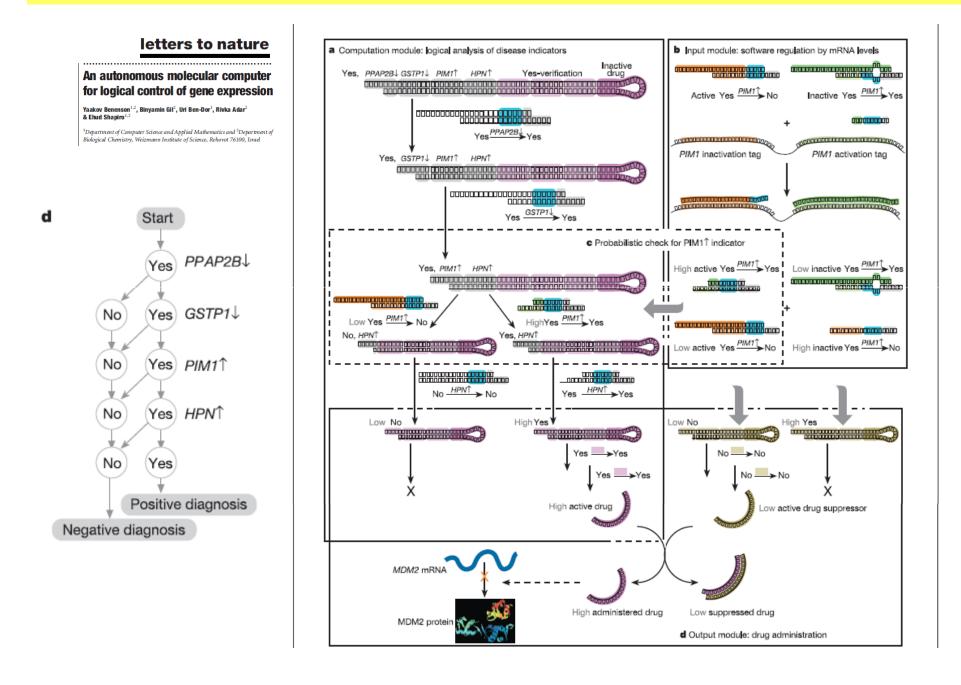
-self-assembly-6 nm feature spacing-versatile template / etch mask

# **DNA as a Computational Material**

## **Aptamers (Sensors)**



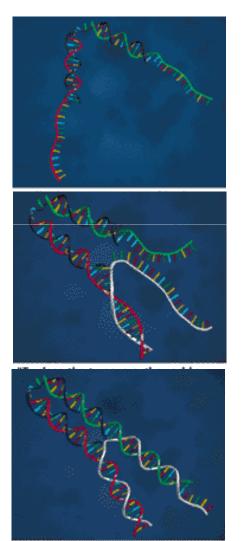
### **Computation:** Curing Cancer with one AND Gate



### **Actuators**

#### **DNA Tweezers**

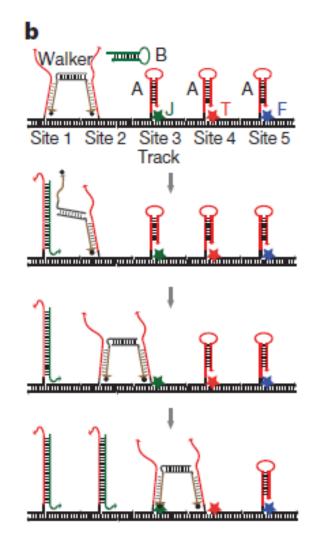
(Yurke & Turberfield, Nature 2000)



"The fuel strand attaches to the handles and draws the two arms of the tweezers together."

#### **DNA Walkers**

(Yin, Choi, Calvert & Pierce, Nature 2008)



## **Summary**

- DNA technology is making great progress
  - $\circ~$  Developing sensor, actuators, and building materials.
  - $_{\odot}~$  All thanks to the programmable nature of DNA.
- DNA computation has also been investigate deeply.
  - DNA tiling systems are Turing complete. They can be used to build 'carpets' with predetermined size and organization.
  - Automata and Turing machines have been demonstrated or designed.
- But there is still space for creativity
  - $\circ~$  What is the 'best' way to write algorithms with DNA?
- What is DNA nanotech for? Ultimately:
  - $\circ~$  To construct 'arbitrary' nanomaterials.
  - $\circ~$  To compute 'in vivo'.

# Computation by DNA Strand Displacement

## **DNA Computing**

#### • Early DNA Computing

- Demonstrated computation by DNA hybridization [Adelman].
- Why DNA? Widely available mature technology.
- Massively concurrent (but still not enough for NP-complete problems).
- Slow and awkward (manual cycling).

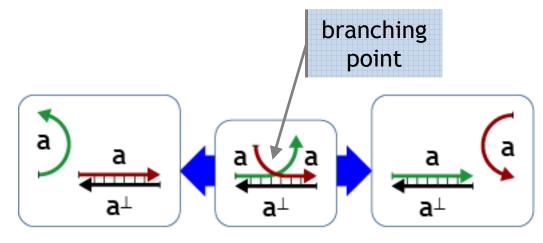
#### New Focus

- $\circ$  Not going to compete with Intel in speed (hours ... days).
- $\circ~$  But can interface with biological systems!
- $\circ~$  For detection and intervention in live organisms.

#### • New Paradigm

- Autonomous DNA computation (mix-and-go) [Yurke&Mills].
- Output readout by fluorescence or atomic microscopy, in vitro.
- Or by influencing cellular mechanisms in vivo [Shapiro survey].

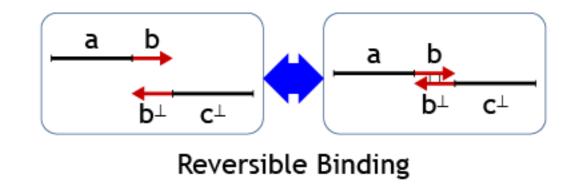
## **Branch Migration**

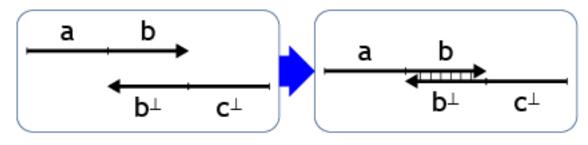


**Branch Migration** 

The branching point moves left and right by a random walk. Until it reaches an end point.

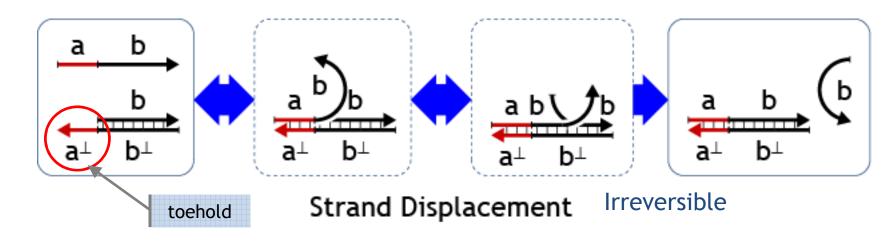
### Short and Long Segments

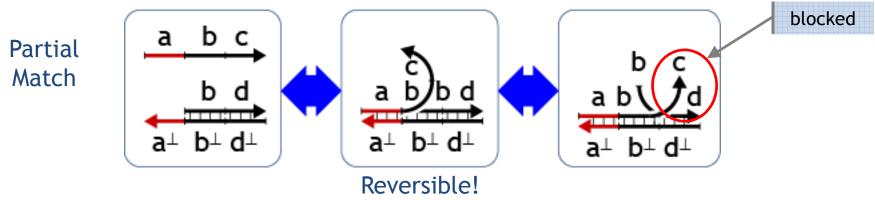




**Irreversible Binding** 

### **Strand Displacement Reaction**

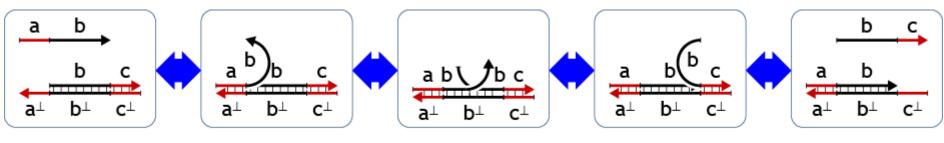




because the random walk is 'reflected' by the blockage

Irreversible match is determined by the toehold **plus** the branch migration region. That is, the toehold is a *cache* for the full address. The toehold must be short enough to guarantee reversible binding, but the branch migration region is practically unlimited. This means that the address space is unlimited.

## **Toehold Exchange Reaction**



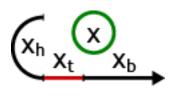
Toehold Exchange

Reversible

## **Signal Strand**

D. Soloveichik, G. Seelig, E. Winfree. **DNA as a Universal Substrate for Chemical Kinetics.** Proc. DNA14.

(We work with a simpler version of their signal stands.)



 $x_{h} = history$  $x_{t} = toehold$  $x_{h} = binding$ 

The history  $x_h$  is not part of signal recognition: strands with different histories should behave the same. Hence, x denotes an equivalence class of strands with different histories.

The combination  $x_t, x_b$  identifies the signal x.

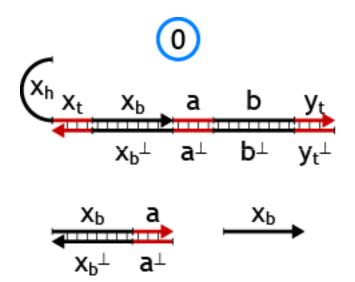
If  $x \neq y$  then x and  $y^{\perp}$  are not supposed to hybridize.

## **Signals and Gates**

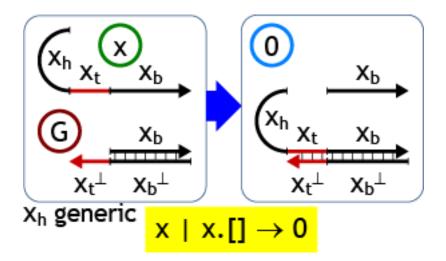
- Signals "x" are always positive strands
- Gates "x.y" always have a negative strand toehold and backbone.
  - that is, the input "x" is implicitly perp'ed
  - and the output "y" is another positive signal
- This separation helps the DNA realization, as one can use 3-letter alphabets (ATC/ATG) for each strand, minimizing secondary structure and entanglement.

### **Inert Systems**

A system is considered *inert* (terminated) if it has no free toeholds.



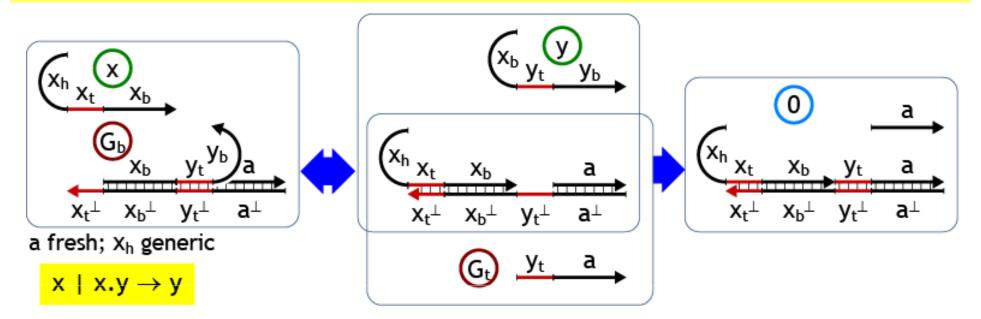
## x.[] Annihilator Gate



This is just the strand displacement reaction, but seen as a gate absorbing a signal x and producing nothing (0 = inert).

Any history segment that is not determined by the gate structure is said to be 'generic' (can be anything).

## x.y Transducer Gate

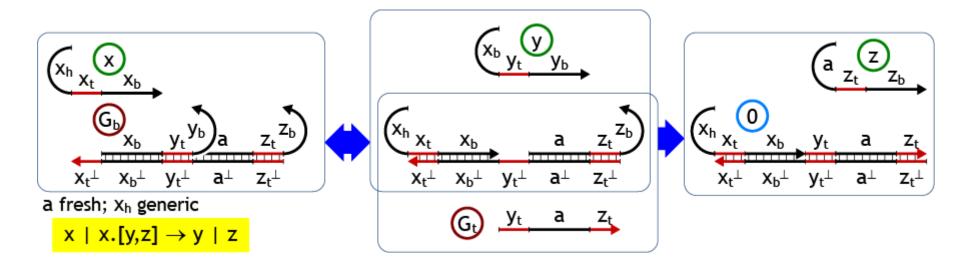


 $G_b, G_t$  (gate backbone and trigger) form the transducer.

Any history segment that is not determined by the gate structure is said to be 'generic' (can be anything).

Any gate segment that is not a non-history segment of an input or output signal is taken to be 'fresh' (globally unique for the gate), to avoid possible interferences.

## x.[y,z] Fork Gate

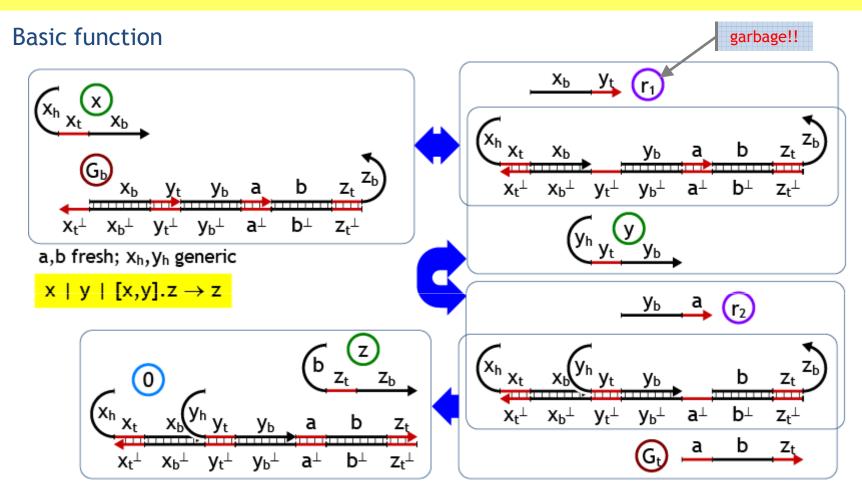


 $G_b, G_t$  (gate backbone and trigger) form the transducer.

Any history segment that is not determined by the gate structure is said to be 'generic' (can be anything).

Any gate segment that is not a non-history segment of an input or output signal is taken to be 'fresh' (globally unique for the gate), to avoid possible interferences.

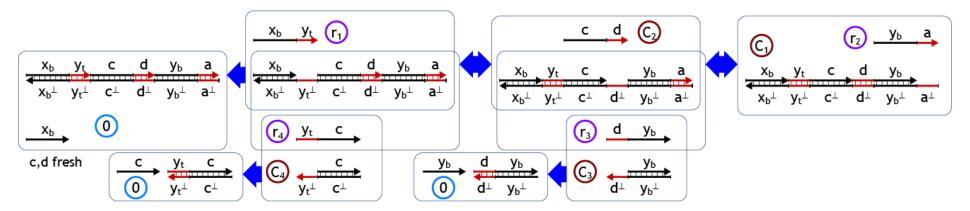
## [x,y].z Join Gate (function)



Join can be implemented by a 'reversible-AND gate' taking two sequential inputs where the first one is reversible (Soloveichik Fig.3), so that x is not actually absorbed until y is found. The 'garbage'  $r_1$  must not be collected until y is found: this is signaled by the release of  $r_2$ .

## [x,y].z Join Gate (collection)

#### Garbage Collection

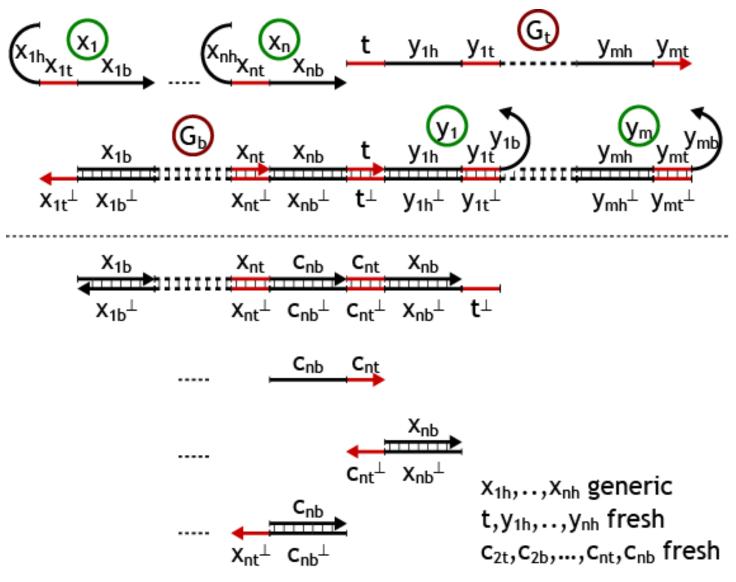


Garbage collection of  $r_1$  is needed for join to work well. This is done by another reversible-AND between  $r_1$  and  $r_2$ , triggered by the release of  $r_2$ . This second reversible-AND leaves garbage too  $(r_3, r_4)$ , but this can be collected immediately, as we know by construction that both inputs  $r_1, r_2$  are available and we need not wait to revert their bindings.

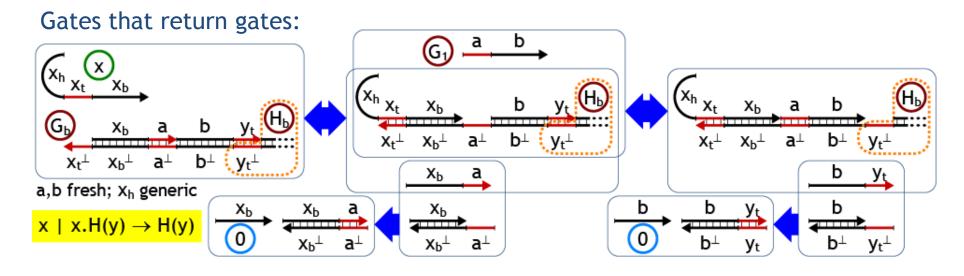
The extra intermediate c,d segments separate the  $r_1$  binding from the  $r_2$  binding. Without them, a segment  $y_t:y_b$  (instead of  $y_t:c$  and  $d:y_b$ ) would be released: that is y!

## [x<sub>1</sub>,..,x<sub>n</sub>].[y<sub>1</sub>,..,y<sub>m</sub>] General Join/Fork Gate

 $x_1 | ... | x_n | [x_1,...,x_n].[y_1,...,y_m] \rightarrow y_1 | ... | y_m$ 

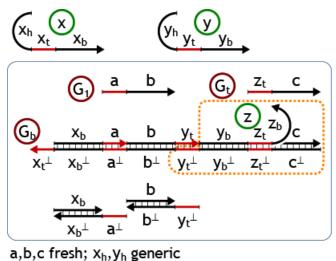


### x.H(y) Curried Gates



For example, x.y.z:

 $x \mid x.y.z \rightarrow y.z$ 

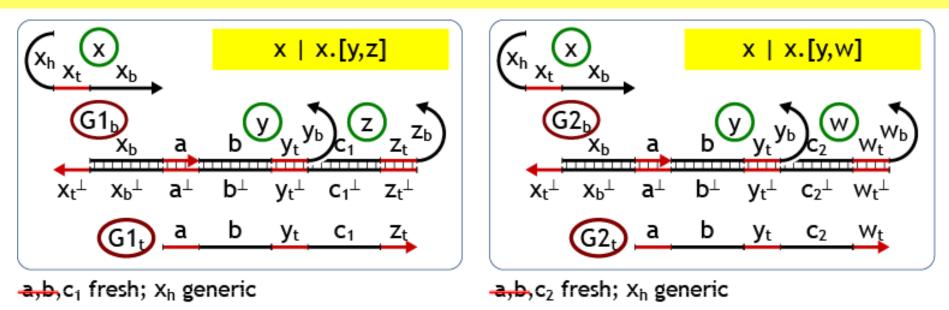


This means we can have gates of the form:

G ::= 
$$[x_1,...,x_n]$$
. $[x'_1,...,x'_m]$  :  
 $[x_1,...,x_n]$ .G  
n≥1, m≥0

## Design, Compilation and Verification Challenges

### Exercise 3: x.[y,z] | x.[y,w] Interference



• Suppose we 'forgot' to take a,b fresh, so they are shared by the two gates. Something goes horribly wrong from these initial conditions:

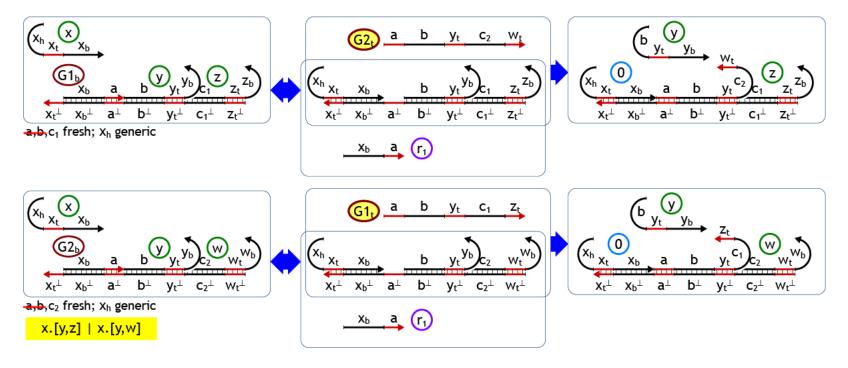
x | x.[y,z] | x | x.[y,w]

```
where x.[y,z] = G1_b, G1_t and x.[y,w] = G2_b, G2_t
```

• What goes wrong?

#### **Exercise 3 Solution**

Deadlocks! Consider x | x | x.[y,z] | x.[y,w], and suppose we had taken c fresh (hence different  $c_1, c_2$ ), but did *not* used gate-unique segments for a,b:



The  $G2_t$  trigger can bind to the wrong  $G1_b$  backbone and get stuck there, and vice versa, without ever releasing z or w.

This is just a made-up problem, but one must watch out for all kinds of possible interferences.

## Exercise 4: x.y.z | [x,y].w Interference

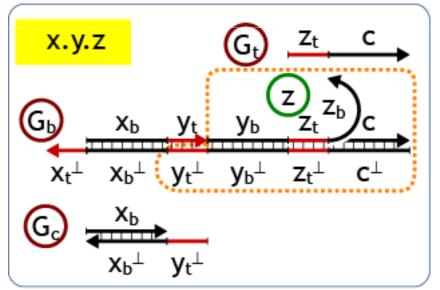
Consider curried gates without the a,b segments (example below): instead of releasing  $x_b$ , a and b,  $y_t$  segments, they would release  $x_b$ ,  $y_t$ .

But that is exactly the strand  $r_1$  of an [x,y].w gate: the strand that reverts the x input. This definitely causes an interference between x.y.z and [x,y].w.

Find a situation where the presence (x.y.z as below) or absence (x.y.z as in previous slide) of this interference causes different outcomes.

Hint: it changes outcome probability.

Note: the a,b segments prevent the interference.



c fresh; X<sub>h</sub>, y<sub>h</sub> generic (without the a,b segments)

### Addressing the Challenges

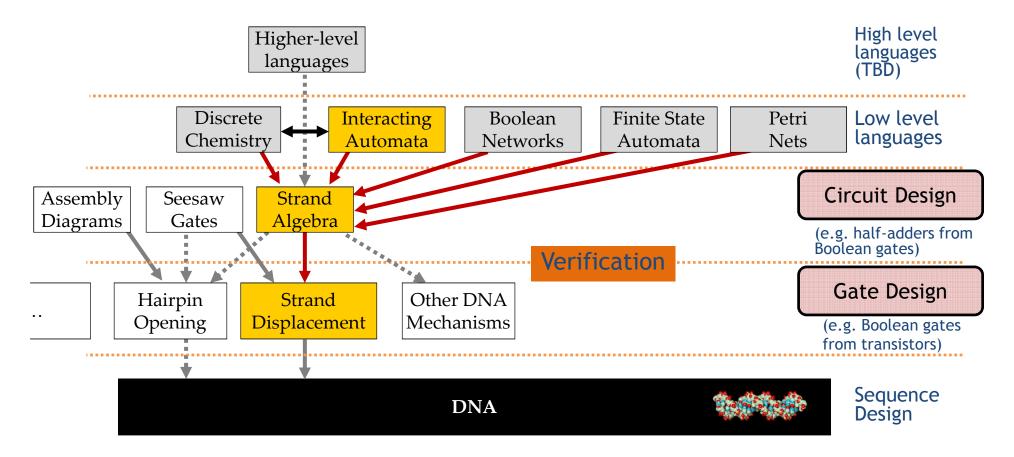
- We need to formally specify
  - $\circ~$  The intended behavior of DNA gates.
  - $\circ$  Their implementation.

#### • We need to verify

- $\circ~$  That the implementation satisfies the specification.
- $\circ~$  In all possible 'soups' (contexts).
- Possibly by modelchecking (the state space is highly combinatorial).

### **DNA Compilation**

#### Separating Circuit Design from Gate Design



### Summary

- DNA strand displacement technology
  - $\circ$  Provides a way of implementing abstract signal transducer networks.
  - Fork gates and Join gates are the main components.
- How powerful it this style of computation?
- How do we verify its correctness?

## **Strand Algebra**

### **Strand Algebra**

Ρ	::=	х÷	$[x_1,,x_n].[y_1,,y_m]$	:	0 :	P P :	P*	n≥1, m≥0
---	-----	----	-------------------------	---	-----	-------	----	----------

X	is a signal
$[x_1,, x_n] \cdot [y_1,, y_m]$	is a <i>gate</i>
0	is an <i>inert solution</i>
PIP	is <i>parallel composition</i> of signals and gates
P*	is a <i>population</i> (multiset) of signals and gates

**Reaction Rule** 

$$x_1 \mid \ldots \mid x_n \mid [x_1, \ldots, x_n] . [y_1, \ldots, y_m] \rightarrow y_1 \mid \ldots \mid y_m$$

Auxiliary rules (axioms of diluted well-mixed solutions)

 $\begin{array}{ll} \mathsf{P} \to \mathsf{P'} & \Rightarrow & \mathsf{P} \mid \mathsf{P''} \to & \mathsf{P'} \mid \mathsf{P''} & & \mathsf{Dilution} \\ \mathsf{P} \equiv \mathsf{P}_1, \, \mathsf{P}_1 \to \mathsf{P}_2, \, \mathsf{P}_2 \equiv \mathsf{P'} & \Rightarrow & \mathsf{P} \to \mathsf{P'} & & \mathsf{Well Mixing} \end{array}$ 

Where  $\equiv$  is a congruence relation (syntactical 'chemical mixing') with  $P^* \equiv P \mid P^*$  for unbounded populations.

### **Compiling Strand Algebra to DNA**

P ::= x :  $[x_1,..,x_n]$ . $[y_1,..,y_m]$  : 0 : P|P : P\* n≥1, m≥0

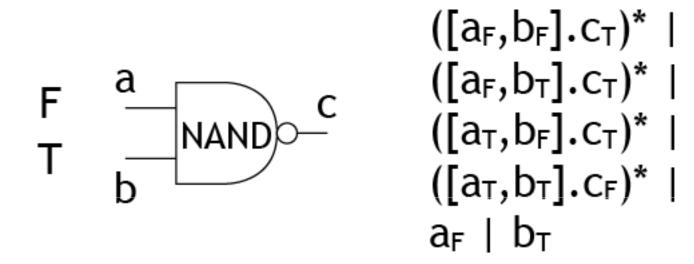
- compile(x) =  $(x_h, x_t, x_b)$
- compile([x<sub>1</sub>,..,x<sub>n</sub>].[y<sub>1</sub>,..,y<sub>m</sub>]) =

$$= \underbrace{\begin{array}{c} t \\ x_{1h} \\ x_{1b} \\ x_{1t} \\ x_{1b} \\ x_{1t} \\ x_{1b} \\ x_{1t} \\ x_{1b} \\ x_{nt} \\ x_{nt}$$

- compile(0) = empty solution
- o compile(P | P') = mix(compile(P), compile(P'))
- compile(P\*) = population(compile(P))

#### **Boolean Networks**

Boolean Networks to Strand Algebra



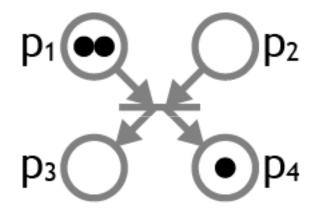
This encoding is *compositional*, and can encode *any* Boolean network:

- multi-stage networks can be assembled (combinatorial logic)
- network loops are allowed (sequential logic)

#### **Petri Nets**

Petri Nets to Strand Algebra

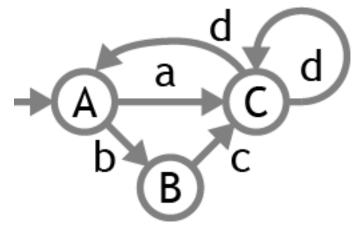
Transitions as Gates Place markings as Signals



### 

#### **Finite State Automata**

#### FSA to Strand Algebra

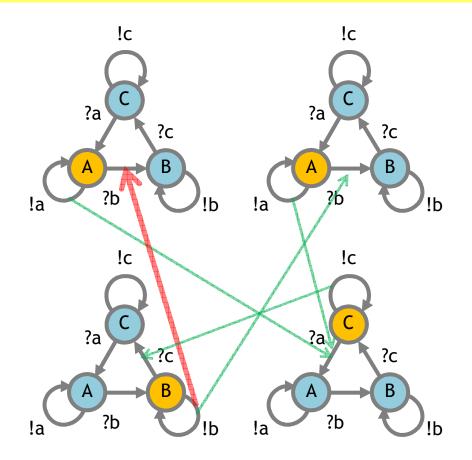


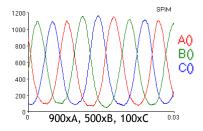
Input strings

a,b,c,d

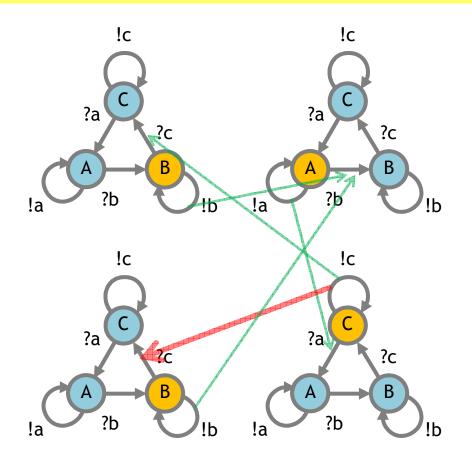
([A,a].[C,τ])\* | ([A,b].[B,τ])\* | ([B,c].[C,τ])\* | ([C,d].[C,τ])\* | ([C,d].[A,τ])\* | ΑΙτ τ.[**a**,σ<sub>1</sub>]|

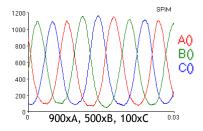
 $[\sigma_1, \tau].[b, \sigma_2]|$  $[\sigma_2, \tau].[c, \sigma_3]|$  $[\sigma_3, \tau]. d$ 



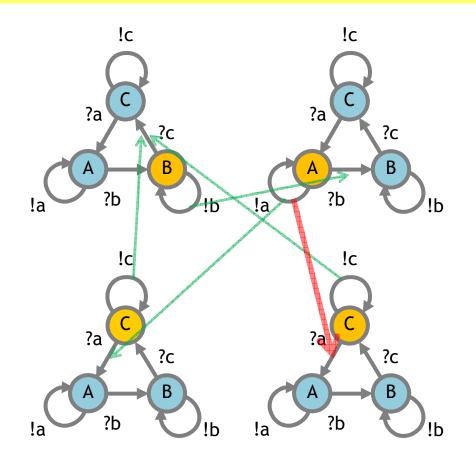


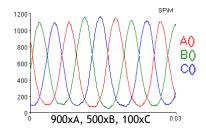
([A,B].[B,B])\* | ([B,C].[C,C])\* | ([C,A].[A,A])\* | **A | A | B | C** 



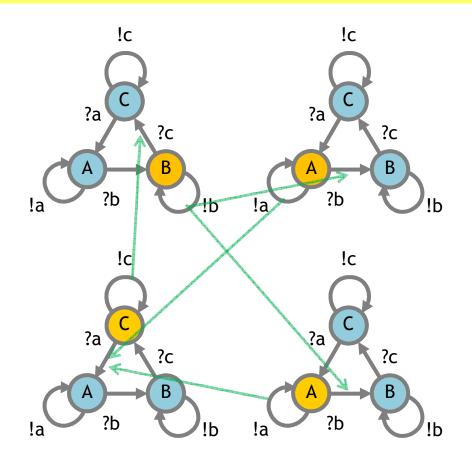


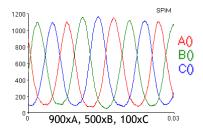
([A,B].[B,B])\* | ([B,C].[C,C])\* | ([C,A].[A,A])\* | A | B | B | C





([A,B].[B,B])\* | ([B,C].[C,C])\* | ([C,A].[A,A])\* | A | B | C | C

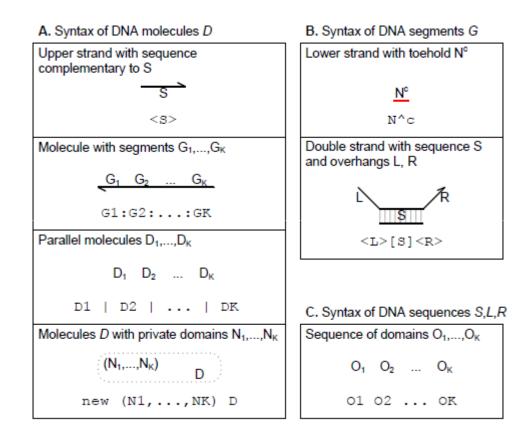




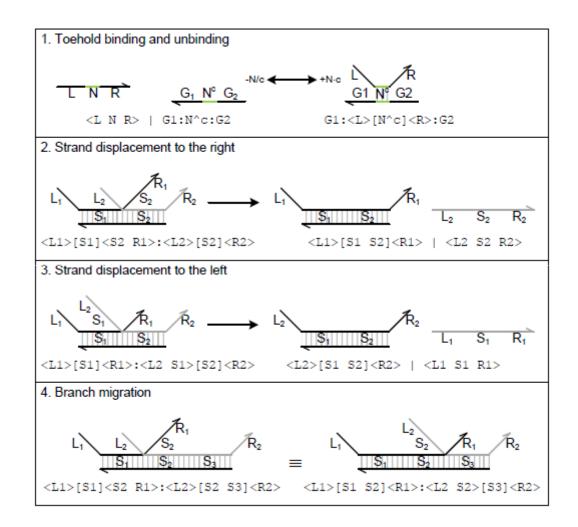
([A,B].[B,B])\* | ([B,C].[C,C])\* | ([C,A].[A,A])\* | A | A | B | C

## Strand Displacement Intermediate Language

### Syntax

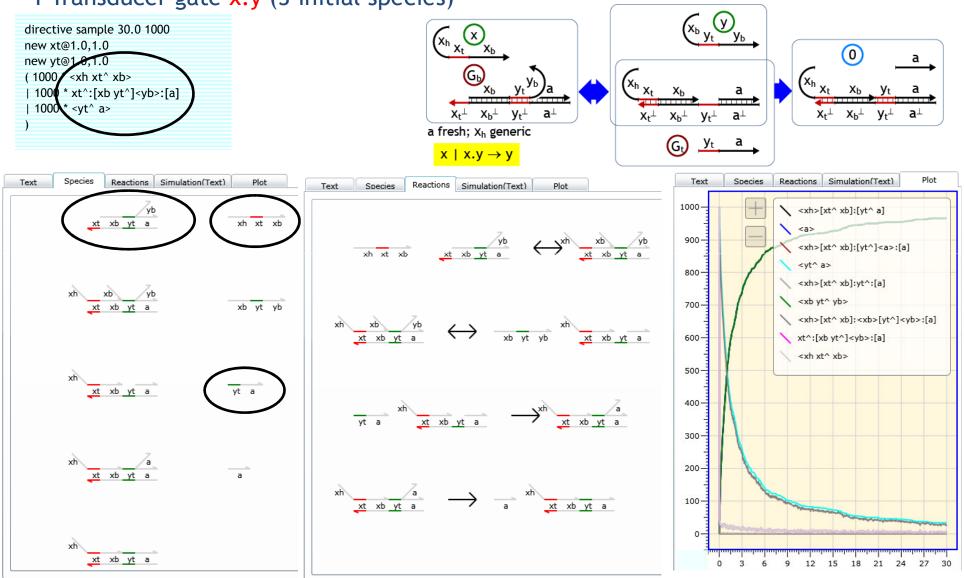


### **Dynamics**



### **Strand Displacement Simulation Tool**

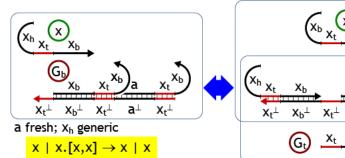
#### 1 Transducer gate x.y (3 initial species)

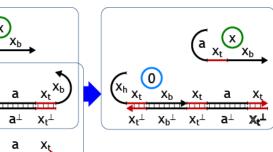


#### **Strand Displacement Simulation Tool**

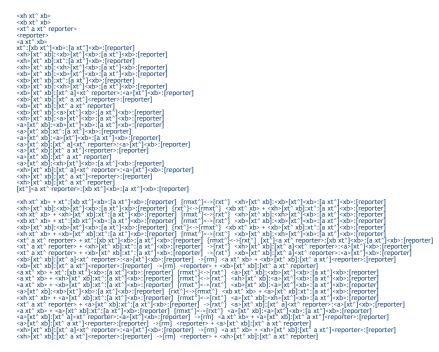
#### Fork Chain Reaction x.[x,x] (3 initial species)

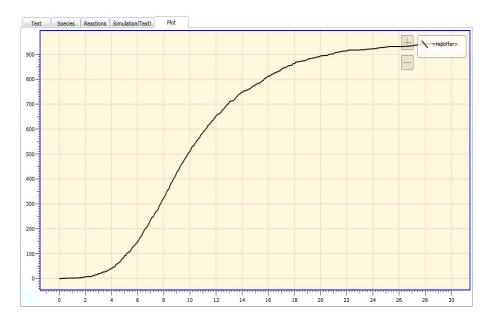
directive sample 30.0 1000 directive plot "<reporter>" new xt@ 1.0, 1.0 (1 \* <xh xt^ xb> | 1000 \* xt^:[xb xt^]<xb>:[a xt^]<xb>:[reporter] | 1000 \* <xt<sup>^</sup> a xt<sup>^</sup> reporter>





#### 28 Species, 22 Reactions





х

а

a⊥

Xt⊥

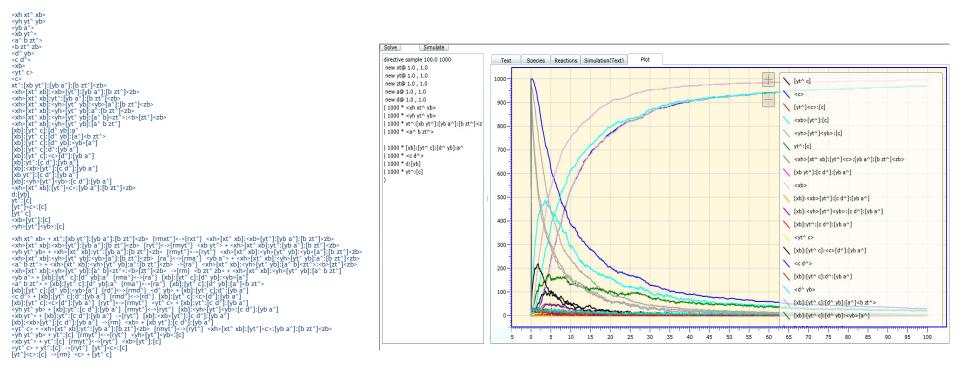
xt

### **Strand Displacement Simulation Tool**

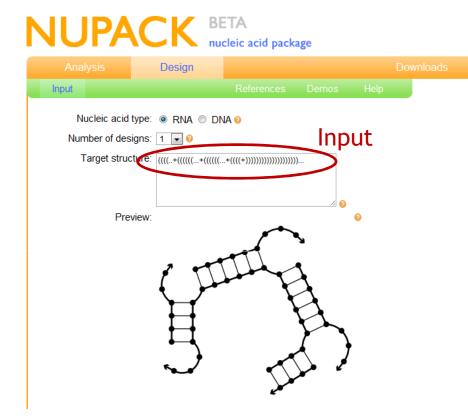
#### 1 Join gate with garbage collection [x,y].z (8 initial species)

directive sample 1000.0 1000 new xt@ 1.0, 1.0 new yt@ 1.0, 1.0 new z@ 1.0, 1.0 new a@ 1.0, 1.0 new d@ 1.0, 1.0 (1000 * <xh xb="" xt^="">   1000 * <yh yb="" yt'="">   1000 * xt^;kb yt^]:[b a^]:[b zt^]<zb>   1000 * <a^ b="" zt^="">   1000 * (xb]:[yt^ c]:[d^ yb]:a^</a^></zb></yh></xh>	$(\overbrace{x_{b}}^{(k)}, \overbrace{x_{b}}^{(k)}, \overbrace{y_{b}}^{(k)}, \overbrace{y_{b}}^{(k)}, \overbrace{z_{b}}^{(k)}, \overbrace{y_{b}}^{(k)}, \overbrace{z_{b}}^{(k)}, \overbrace{z_{b}}^{($	$\begin{array}{c c} x_{b} & y_{b} \\ \hline \\ $	$\begin{array}{c c} x_{b} & y_{b} & c & d & y_{b} & a \\ \hline x_{b} & y_{b}^{\perp} & c^{\perp} & d^{\perp} & y_{b}^{\perp} & a^{\perp} \\ \hline x_{b} & y_{b}^{\perp} & c^{\perp} & d^{\perp} & y_{b}^{\perp} & a^{\perp} \\ \hline x_{b} & y_{b}^{\perp} & c^{\perp} & d^{\perp} & y_{b}^{\perp} & a^{\perp} \\ \hline x_{b} & y_{c}^{\perp} & c^{\perp} & d^{\perp} & y_{b}^{\perp} & a^{\perp} \\ \hline x_{b} & y_{c} & c^{\perp} & d^{\perp} & y_{b}^{\perp} & a^{\perp} \\ \hline x_{b} & y_{c} & c^{\perp} & d^{\perp} & y_{b}^{\perp} & a^{\perp} \\ \hline x_{b} & y_{c} & c^{\perp} & c^{\perp} & d^{\perp} & y_{b}^{\perp} & a^{\perp} \\ \hline x_{b} & y_{c} & c^{\perp} & c^{\perp} & c^{\perp} & c^{\perp} & d^{\perp} & y_{b}^{\perp} & a^{\perp} \\ \hline x_{b} & y_{c} & y_{b} & d^{\perp} & y_{b}^{\perp} & d^{\perp} \\ \hline x_{b} & y_{c} & c^{\perp} & c^{\perp} & c^{\perp} & c^{\perp} & d^{\perp} & y_{b}^{\perp} & a^{\perp} \\ \hline x_{b} & y_{c} & c^{\perp} & c^{\perp} & c^{\perp} & c^{\perp} & d^{\perp} & y_{b}^{\perp} & a^{\perp} \\ \hline x_{b} & y_{c} & z^{\perp} & c^{\perp} & c^{\perp} & c^{\perp} & d^{\perp} & y_{b}^{\perp} & a^{\perp} \\ \hline x_{b} & y_{b} & x_{b} & y_{b} & x_{b} & y_{b} & d^{\perp} & y_{b}^{\perp} \\ \hline x_{b} & y_{c} & z^{\perp} & c^{\perp} & c^{\perp} & c^{\perp} & d^{\perp} & y_{b}^{\perp} \\ \hline x_{b} & y_{b} & x_{b} & y_{b} & x_{b} & y_{b} & x_{b} & y_{b} & x_{b} \\ \hline x_{b} & y_{c} & z^{\perp} & z^{\perp} & z^{\perp} & z^{\perp} \\ \hline x_{b} & y_{b} & z^{\perp} & z^{\perp} & z^{\perp} & z^{\perp} \\ \hline x_{b} & y_{b} & z^{\perp} & z^{\perp} & z^{\perp} & z^{\perp} \\ \hline x_{b} & y_{b} & z^{\perp} & z^{\perp} & z^{\perp} \\ \hline x_{b} & y_{b} & z^{\perp} & z^{\perp} & z^{\perp} \\ \hline x_{b} & y_{b} & z^{\perp} & z^{\perp} & z^{\perp} \\ \hline x_{b} & y_{b} & z^{\perp} & z^{\perp} & z^{\perp} \\ \hline x_{b} & z^{\perp} & z^{\perp} & z^{\perp} & z^{\perp} & z^{\perp} \\ \hline x_{b} & z^{\perp} & z^{\perp} & z^{\perp} & z^{\perp} & z^{\perp} & z^{\perp} \\ \hline x_{b} & z^{\perp} & z^{\perp} & z^{\perp} & z^{\perp} & z^{\perp} \\ \hline x_{b} & z^{\perp} \\ \hline x_{b} & z^{\perp} \\ \hline x_{b} & z^{\perp} \\ \hline x_{b} & z^{\perp} &$
1000 * cd <sup>+</sup> > (c]   1000 * d:[yb]   1000 * yt <sup>+</sup> :[c] )	$\underbrace{ \begin{pmatrix} 0 & \begin{pmatrix} b & \underline{c} \\ z_t & z_b \end{pmatrix} \\ (x_h x_t & x_b y_h y_t & y_b & a & b & z_t \\ x_{\perp}^{\perp} & x_b^{\perp} & y_t^{\perp} & y_b^{\perp} & a^{\perp} & b^{\perp} & z_{\perp}^{\perp} \end{pmatrix} }_{\mathbf{X}_{\perp}^{\perp} \mathbf{X}_{\perp}^{\perp} \mathbf{X}_$	$(x_{h} \xrightarrow{x_{h}} x_{h} \xrightarrow{y_{h}} y_{t} \xrightarrow{y_{h}} \xrightarrow{b} \xrightarrow{z_{h}} x_{h} \xrightarrow{z_{h}} y_{t} \xrightarrow{y_{h}} y_{h} \xrightarrow{a^{\perp}} a^{\perp} \xrightarrow{b^{\perp}} z_{t}^{\perp}$ $(c_{h} \xrightarrow{a} \xrightarrow{b} z_{t}$	

#### 34 Species, 19 Reactions



#### **Sequence Design**



#### NUPACK BETA nucleic acid package

Analysis		De	sign				
Input	Results			References	Demos	Help	
	summary	0					
equence de	signs 📀						
equence de Average percentag of correct nucleotide ©	Avera e numl incor	per of	GC content	Sequence Ø	(	Dutp	out

Copyright © 2007-2009 Caltech. All rights reserved. Contact Funding Terms of use

# Conclusions

### Conclusion

#### • Nucleic Acids

• Programmable matter

#### • DNA Strand Displacement

 $\circ~$  A computational mechanism at the molecular level

#### • DNA Compilation

- High-level languages (Boolean Networks, Petri Nets, Interacting Automata)
- o Intermediate languages.
- Sequence generation.

#### • Tools

- $\circ$  Thermodynamic analysis.
- $\circ~$  Simulation.
- $\circ~$  Verification (not yet).

### Abstract

Nucleic acids (DNA/RNA) encode information digitally, and are currently the only truly 'userprogrammable' entities at the molecular scale. They can be used to manufacture nano-scale structures, produce physical forces, act as sensors and actuators, and do computation in between. Eventually we will be able to interface then with biological machinery to detect and cure diseases at the cellular level under program control.

The technology to create and manipulate them has existed for many years, but the imagination necessary to exploit them has been evolving slowly. Recently, some very simple computational schemes have been developed that are autonomous (run on their own once started) and involve only short (easily synthesizable) DNA strands with no other complex molecules. To get this started, one emails some short character strings to a company to get the DNA strands built, mixes them up, and reads the output (fluorescence) with a camera. And yes, this can be done in your kitchen, more or less.

But of course we need programming tools. Molecular design is required to produce molecules that fold, or do not fold, or stick, or do not stick in the desired ways: this can be achieved fairly predictably only for DNA/RNA (e.g. not for proteins). On that basis one can design various kinds of 'logic gates' and 'computational architectures', which is where much of the imagination is currently needed.

Then one needs programming languages both at the level of gate implementation (where Andrew Phillips in Cambridge is building a strand-level language and simulator), and at the level of circuit implementation (where I will describe a Strand Algebra for implementing e.g. automata and Petri nets). Since DNA computation is massively concurrent, some tricky and yet familiar issues arise, like having to formally verify gate designs to avoid subtle deadlocks and race conditions, and having to design high-level languages that exploit concurrency and stochasticity.